

# The potential of Lantana camara Linn. as a source of quercetin, gallic acid, and tannic acid

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## The potential of *Lantana camara* Linn. as a source of quercetin, gallic acid, and tannic acid

**Abstract:** *L. camara* Linn. as a plant that is invasive and considered a dangerous plant, but contains active substances that are beneficial to health. Active substances contained in the leaves of *L. camara* Linn. including flavonoids, gallic acid, and tannic acid. The purpose of this study was to explore the content of quercetin, gallic acid, and tannic acid in *L. camara* Linn. leaf extract. The methods of this study including leaf extract of *L. camara* Linn. were tested organoleptic, pH, quercetin equivalent flavonoid (QEF), gallic acid equivalent phenolic (GAEP), and tannic acid equivalent tannin (TAET). Measurement of quercetin equivalent flavonoid (QEF), gallic acid equivalent phenolic (GAEP), and tannic acid equivalent tannin (TAET) levels was carried out with a spectrophotometer. The QEF content of *L. camara* Linn. leaf extract is  $0.428 \pm 0.004$  mg/g. The GAEP content of *L. camara* Linn. leaf extract is  $0.288 \pm 0.002$  mg/g, while the content of TAET is  $0.384 \pm 0.009$  mg/g. This study confirmed the presence of flavonoids, phenols, and tannins in *L. camara* Linn. leaf extract, either extracted with ethanol or with other solvents, such as acetone or petroleum ether. The novelty of this study that the variations in active substances levels, that is can be used as an option in the exploration and utilization of *L. camara* Linn. Thus, *L. camara* Linn., not only considered as a wild plant that endangers the environment, but can be used as a source for exploration of QEF, GAEP, and TAET.

24

**Keywords:** *Lantana camara* Linn., organoleptic, quercetin equivalent flavonoid, gallic acid equivalent phenolic, tannic acid equivalent tannin

## 马缨丹的潜力。作为槲皮素、没食子酸和单宁酸的来源

抽象的

*L. camara* Linn. 作为一种侵入性植物，<sup>19</sup> <sup>19</sup> <sup>25</sup> <sup>26</sup> <sup>27</sup> 被认为是危险植物，但含有对健康有益的活性物质。*L. camara* Linn 叶子中含有的活性物质。包括类黄酮、没食子酸和鞣酸。本研究的目的是探讨 *L. camara* Linn 叶提取物中槲皮素、没食子酸和单宁酸的含量。本研究的方法包括 *L. camara* Linn 的叶提取物。测试了感官、pH、槲皮素等效类黄酮 (QEF)、没食子酸等效酚类 (GAEP) 和单宁酸等效单宁 (TAET)。使用分光光度计测量槲皮素等效类黄酮 (QEF)、没食子酸等效酚类 (GAEP) 和单宁酸等效单宁 (TAET) 水平。*L. camara* Linn 的 QEF 含量。叶提取物为  $0.428 \pm 0.004$  mg/g。*L. camara* Linn 的 GAEP 内容。叶提取物为  $0.288 \pm 0.002$  mg/g，而 TAET 的含量为  $0.384 \pm 0.009$  mg/g。该研究证实了 <sup>2</sup> *L. camara* Linn 中存在类黄酮、酚类和单宁酸。叶提取物，用乙醇或其他溶剂（如丙酮或石油醚）提取。本研究的新颖之处在于活性物质水平的变化，即可以作为探索和利用 *L. camara* Linn 的一种选择。因此，*L. camara* Linn. 不仅被认为是一种危害环境的野生植物，而且可以作为探索 QEF、GAEP 和 TAET 的资源。

关键词: *Lantana camara* Linn., 感官, 槲皮素等效类黄酮, 没食子酸等效酚类, 单宁酸等效单宁

## 1. Introduction

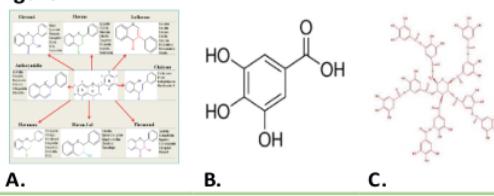
Flavonoids are polyphenolic compounds, found in various parts of plants. There are 8 classes of flavonoids, namely: flavones, flavonols, flavanones, flavanonol, isoflavones, flavantriol, anthocyanidins, and chalcone [1]. The benefits of flavonoids in health include being anti-cancer, anti-oxidative, anti-inflammatory, stimulating bone formation [2]. Recent studies have shown that flavonoids have antiviral activity against SARS-CoV-2 [3]. Gallic acid is a phenol compound, and is known by another name, 3,4,5-trihydroxybenzoic acid. Gallic acid has the chemical formula (chemical structure)  $C_6H_2(OH)_3COOH$  [4]. The results of recent studies demonstrate that gallic acid is produced by *Swietenia macrophylla* [5]. In general, plants produce gallic acid [6]. The benefits of gallic acid in the health sector include anti-microbial, prooxidant, anti-oxidant, anti-inflammatory, anti-platelet, anti-dengue, anti-cancer, and anti-apoptotic [7]. Tannins are phenolic compounds found in plants. There are 2 groups of tannins, namely hydrolysable and condensed tannins. Gallotannins are examples of hydrolysable tannins, while catechins and gallic acid are examples of condensed tannins [8]. As is the case with flavonoids and gallic acid, tannins are also produced by plants, i.e. *Hibiscus sabdariffa* tea [9], *Dimocarpus longan* [10]. The biological activities of tannins include antimicrobial, antidiabetic, antioxidant, and cardioprotective [11].

The results of previous studies showed the content of flavonoids, gallic acid, and tannins in the following types of plants. QEF levels in methanol extract of *Melastoma malabathricum* L. fruit are 6,827 mg/g, while GAEP levels are 154,880 mg/g extract [12]. In addition, it has also been reported that stem bark extract from *M. gigantea* contains flavonoids 25.2 mg/g [13]. It is interesting to note that the content of phenolics catechins is equivalent in various varieties of *Vitis* sp. classified as quite high (>900 mg/L) [14]. The results of other studies showed that various extraction methods against *M. malabathricum* L. shows variation in GAEP levels [15]. The results of this study are in line with the results of research demonstrating that tannin content is different in various cultivars of *Vitis* species Red Wines measured by various measurement methods [14].

Previously [20] we measured QEF levels at various concentrations of *L. camara* Linn. leaf extract cream. *L. camara* Linn. leaf collection was obtained from the area of Tanjakan Cino Mati, Pleret District, Bantul Regency, Special Region of Yogyakarta, Indonesia [16]. *Lantana camara* Linn. is an invasive plant [17], so it is

considered a dangerous plant in Indonesia [18]. Several researchers in Indonesia have explored the active ingredients of *L. camara* Linn. to be used in the field of Health [19, 20]. One possible use of *L. camara* Linn. in the field of health, namely utilizing the content of active substances including flavonoids, gallic acid, and tannic acid.

Since *L. Camara* Linn. is invasive, and contains active substances that are beneficial to health, we hope that the plant can be used as a source of flavonoids, gallic acid, and tannic acid. Research still needs to be done to explore the content of flavonoids, gallic acid, and tannic acid in *L. camara* Linn. leaf extract. We hope that the results of this study can be used as a reference option about the potential of *L. camara* Linn. as a source of active ingredients in the form of quercetin equivalent flavonoid (QEF), gallic acid equivalent phenolic (GAEP), and tannic acid equivalent tannin (TAET). The structural formulas of flavonoids, gallic acid, and tannic acid are presented in Figure 1.



**Figure 1.** The structural formula of flavonoids, gallic acid, and tannic acid. A. Basic structure and classification of flavonoids [1]. B. Gallic acid (3,4,5-Trihydroxybenzoic acid) [4, 21]. C. Structure of tannic acid [22].

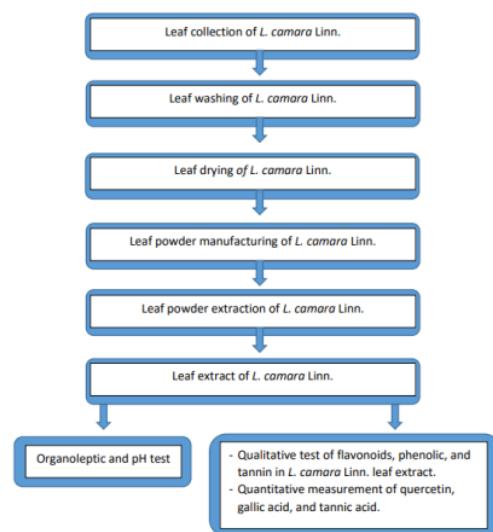
## 2. Material and Methods

### 2.1. Research design

The design of this research is a laboratory experimental research. The main process of these research showed in Figure 2.

### 2.2. Leaves collection of *L. camara* Linn.

Leaves of *L. camara* Linn. collected from Tondano Kamangta Suluan street, Tombulu District, Minahasa Regency, North Sulawesi Province, Indonesia ( $1^{\circ}21'46.6''N$   $124^{\circ}54'13.0''E$ ). The location can be accessed at <http://goo.gl/maps/nc1SVYhFU39q8nMz8>.



**Figure 2.** The main process of these research

The activity was carried out in December 2022. The collected leaves are then washed under running water, then covered with a black cloth, and dried in the hot sun. Leaves of *L. camara* Linn. which had been dried, ground into powder, then sifted to obtain a fine powder. Fine powder of *L. camara* Linn. leaves it is further extracted using 96% ethanol. *L. camara* Linn. leaf extract obtained in a viscous form, dark green in color, then put into sterile bottles, and stored in a refrigerator. The extracts it is ready to be tested.

### 2.3. Organoleptic, and pH test of *L. camara* Linn. leaf extract

Organoleptic tests performed on *L. camara* Linn. leaf extracts include shape, smell, and color. In addition, pH measurements were also carried out on *L. camara* Linn. leaf extracts [23, 24, 25].

### 2.4. Qualitative test of flavonoids, phenolic, and tannin in *L. camara* Linn. leaf extract.

#### 2.4.1. Qualitative test of flavonoids

Fifty mg of sample was dissolved with 5 mL ethanol in a test tube, then heated for five minutes. Next, add a few drops of concentrated HCl, then add 0.2 g of Mg powder. A positive result is indicated by the onset of dark red color for 3 minutes.

#### 2.4.2. Qualitative test of phenolic

One mL of sample is dissolved in a test tube containing methanol, then 5% FeCl<sub>3</sub> is added. A positive result in the presence of phenolic compounds is indicated by a change in color to orange-brown.

#### 2.4.3. Qualitative test of tannin

Fifty mg of sample was put into a test tube, then added ethanol until the sample was submerged, then added 2-3 drops of 1% FeCl<sub>3</sub> solution. A positive result for tannin content is indicated by the formation of a bluish black or green color.

### 2.5. Quantitative measurement of phytochemical in *L. camara* Linn. leaf extract.

#### 2.5.1. Measurement of flavonoid levels

Measurement of QEF levels was carried out with aluminum chloride colorimetric assay [26], using the UV-1800 spectrophotometer (Shimadzu Corp. 00787, Serial No. 116351). The standard curve of QEF is duplicated with concentrations of 2, 4, 6, 8 and 10 µg/mL in 80% methanol solvent. One mL of each series of standard solution plus 4 mL distilled water, then added 0.30 mL 5% NaNO<sub>2</sub>, and homogenized, then allowed to stand for 5 minutes. Next, add 0.3 mL to 10% AlCl<sub>3</sub>, and homogenize using a vortex mixer. After 5 min, plus 2 mL of 1 M NaOH, plus 2.4 mL of distilled water until a total volume of 10 mL. Absorbance readings for blanks and standard solutions at a wavelength of 510 nm. The data obtained were used to create a standard curve of flavonoid quercetin equivalent. To measure QEF levels in the samples, it is done by making a sample solution, namely 1 mL of *L. camara* Linn. leaf extract as a substitute for standard solutions. The sample solution is reacted with the same reagents used in standard curve making, as well as absorbance readings. Calculation of total QEF levels by comparing the absorbance of the sample against the standard quercetin curve, the results are expressed as QEF in mg/g sample.

#### 2.5.2. Measurement of gallic acid levels

Phenolic content measurement was performed with Folin-Ciocalteu assay [26, 27], using the UV-1800 spectrophotometer (Shimadzu Corp. 00787, Serial No. 116351). Standard gallic acid curves are duplicated in volumetric flask. The concentration of gallic acid used is 5, 10, 15, 20, 25 µg/mL each 17.9 mL of distilled water. The blank reagent used is distilled water. 1 mL of Folin-Ciocalteu phenol reagent was added to each of the prepared standard solutions, homogenized, 5 minutes, 22 after added 2 mL of 7% Na<sub>2</sub>CO<sub>3</sub> solution and 3.6 mL distilled water, then incubated for 90 minutes

at room temperature. Absorbance readings with a spectrophotometer at a wavelength of 650 nm. To measure GAEP levels in samples, it is done by making a sample solution, namely 1 mL of *L. camara* Linn. leaf extract as a substitute for standard solutions. The sample solution is reacted with the same reagents used on the standard curve, as well as the absorbance readings. Total GAEP content was expressed as mg/g sample.

### 2.5.3. Measurement of tannic acid levels

Measurement of tannin content was performed with Folin-Ciocalteu assay [28], using the UV-1800 spectrophotometer (Shimadzu Corp. 00787, Serial No. 116351). Standard tannic acid curves are duplicated in volumetric flask. The concentration of tannic acid used is 10, 20, 40, 60, 80 µg/mL, each in 9 mL of distilled water. The blank reagent used is distilled water. One mL of each standard solution is put into a flask container containing 7.5 mL of distilled water. To the flask is added 0.5 mL of Folin Denish reagent, allowed to stand 3 minutes, then added 1 mL of saturated Na<sub>2</sub>CO<sub>3</sub> solution, then incubated for 15 minutes. Absorbance readings with a spectrophotometer at a wavelength of 740 nm. To measure TAET levels in samples, it is done by making a sample solution, which is 1 mL of *L. camara* Linn. leaf extract instead of standard solutions. The sample solution is reacted with the same reagents used on the standard curve, as well as the absorbance readings. Total TAET content was expressed as mg/g sample.

### 2.6. Data Analysis

Descriptive analysis was carried out on phytochemical data of *L. camara* Linn leaf extract, namely QEF, GAEP, and TAET levels. The phytochemical content of *L. camara* Linn. leaf extract presented in table and graphic form using the Microsoft Excel program.

## 3. Results

### 3.1. Leaves of *L. camara* Linn.

Leaves of *L. camara* Linn. collected from Tondano Kamangta Suluan street, Tombulu District, Minahasa Regency, North Sulawesi Province, Indonesia (1°21'46.6"N 124°54'13.0"E) presented in Figure 3.



A.

B.

C.

**Figure 3.** *L. camara* Linn. A. *L. camara* Linn. as a wild plant. B. Habitus of *L. camara* Linn. showing stems, leaves, flowers, and fruit. B. Leaves of *L. camara* Linn. as an extraction material. Photographer by Hosea Jaya Edy, December 20, 2022.

### 3.2. Test results organoleptic, and pH test of *L. camara* Linn. leaf extract

The results of organoleptic test, and pH of *L. camara* Linn. leaf extract is presented in Table 1.

**Tabel 1.** Test results organoleptic, and pH of *L. camara* Linn. leaf extract

Type of test	Results
Organoleptic	
• Shape	Semi solid
• smell	Typical smell of <i>L. camara</i> Linn. leaf extract
• color	slightly blackish green
pH	5

Abbreviation: pH=potential of hydrogen

The results of qualitative examination of the active substance of *L. camara* Linn. leaf extract is presented in Table 2.

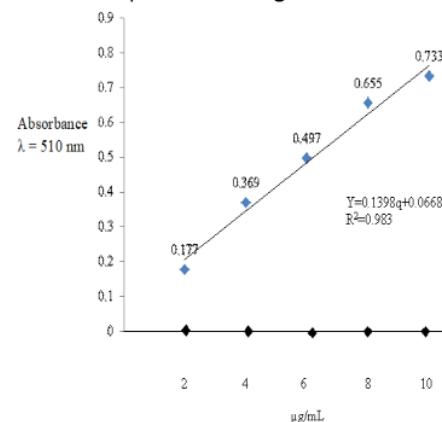
**Tabel 2.** Qualitatively of the active substance of *L. camara* Linn. leaf extract

Compounds tested	Color change results	Result
Flavonoid	Brick red	+
Fenolik	Orange brown	+
Tannin	Greenish-brown transparan	+

Abbreviation: + = positive

### 3.3. Quercetin equivalent flavonoid

The standard curve of quercetin equivalent flavonoid is presented in Figure 4.



**Figure 4.** The standard curve of quercetin equivalent of flavonoid.

The standard curve used for the analysis of QEF levels in this study was  $Y=0.1398q+0.0668$  (Fig. 4). In the equation, Y=absorbance; a=0.1398; b=0.0668; q=quercetin equivalent flavonoid (mg/L) levels. In addition, a standard solution with a concentration of 1.0-10.0  $\mu\text{g/mL}$  obtained a coefficient of determination ( $R^2$ ) =0.983. To calculate total of QEF ( $Q_{\text{QEF}}$ ) per gram of *L. camara* Linn. leaf extract, we used formula  $Q_{\text{QEF}}=q*v*(p/m)$ . In the equation, q=QEF levels in the sample, v=sample volume, p=dilution, and m=sample mass/weight.

QEF levels of *L. camara* Linn. leaf extract presented in Table 3.

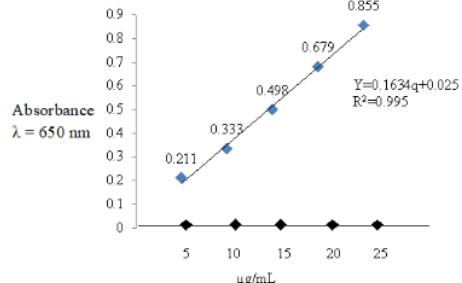
**Table 3.** Quercetin equivalent flavonoid levels of *L. camara* Linn leaf extract.

Sample	Y	a	b	v (L)	p	m (g)	$Q_{\text{QEF}}$ (mg/g)
1	0.661	0.1398	0.0668	0.001	10	0.1	0.425036
2	0.659	0.1398	0.0668	0.001	10	0.1	0.423605
3	0.673	0.1398	0.0668	0.001	10	0.1	0.433319
4	0.669	0.1398	0.0668	0.001	10	0.1	0.431107
5	0.661	0.1398	0.0668	0.001	10	0.1	0.425205
6	0.664	0.1398	0.0668	0.001	10	0.1	0.42712
Mean							0.428
SD							0.004

Abbreviations: Y=absorbance at a wavelength ( $\lambda$ ) 510 nm; a=coefficient; b=constant; v=volume (liters); p=dilution; m=sample weight (gram); q<sub>QEF</sub>=quercetin equivalent flavonoid levels; Q<sub>QEF</sub>=total quercetin equivalent flavonoid; mg/L=milligrams per liter; mg/g=milligrams per gram; SD=standard of deviation.

### 3.4. Phenolic equivalent gallic acid

The standard curve of GAEP is presented in Figure 5.



**Figure 5.** The standard curve of gallic acid equivalent of phenolic

The standard curve of GAEP used in this study is  $Y=0.1634q+0.025$  (Fig. 5). In the equation, Y=absorbance; a=0.1634; b=0.025; q<sub>GAEP</sub>=gallic acid equivalent phenolic (mg/L). In addition, a standard solution with a concentration of 5.0-25.0  $\mu\text{g/mL}$  obtained a coefficient of determination ( $R^2$ ) =0.995. To calculate the amount of GAEP ( $Q_{\text{GAEP}}$ ) per gram of *L. camara* Linn. leaf extract, we used formula  $Q_{\text{GAEP}}=q*v*(p/m)$ . In the equation, q<sub>GAEP</sub>=GAEP levels in the sample, v=sample volume, p=dilution, and m=sample mass/weight.

The results of the analysis of gallic acid equivalent phenolic levels of *L. camara* Linn. leaf extract presented in Table 4.

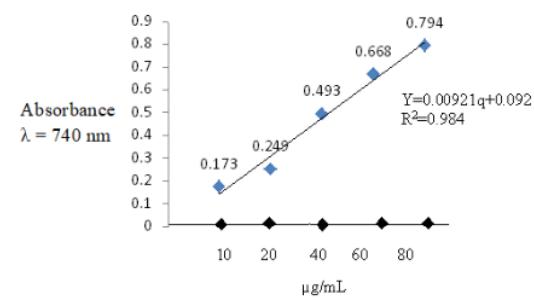
**Table 4.** Levels of gallic acid equivalent phenolic of *L. camara* Linn. leaf extract.

Sample	Y	a	b	v (L)	p	m (g)	$Q_{\text{EGA}}$ (mg/g)
1	0.491	0.1634	0.025	0.001	10	0.1	0.285189
2	0.499	0.1634	0.025	0.001	10	0.1	0.290086
3	0.497	0.1634	0.025	0.001	10	0.1	0.288861
4	0.496	0.1634	0.025	0.001	10	0.1	0.288045
5	0.496	0.1634	0.025	0.001	10	0.1	0.288285
6	0.495	0.1634	0.025	0.001	10	0.1	0.287928
Mean							0.288
SD							0.002

Abbreviations: Y=absorbance at a wavelength ( $\lambda$ ) 650 nm; a=coefficient; b=constant; v=volume (liters); p=dilution; m=sample weight (gram); q<sub>GAEP</sub>=phenolic equivalent to gallic acid levels; Q<sub>EGA</sub>=total gallic acid equivalent phenolic in the sample; mg/L=milligrams per liter; mg/g=milligrams per gram; SD=standard of deviation.

### 3.5. Tannin equivalent tannic acid

The standard curve of TAET is presented in Figure 6.



**Figure 6.** The standard curve of tannin equivalent tannic acid.

The standard curve for TAET used in this study is  $Y=0.00921q+0.092$ . The coefficient of determination ( $R^2$ )

in the equation is 0.984. (Fig. 6). In the equation, Y=absorbance; a=0.1661; b=0.0229; q=TAET levels (mg/L). To calculate the amount of TAET (QTAET) per gram of *L. camara* Linn. leaf extract, we used formula  $Q_{TAET}=q*v*(p/m)$ . In the equation, q=TAET levels in the sample, v=sample volume, p=dilution, and m=sample mass/weight.

Tannic acid equivalent tannin levels in the *L. camara* Linn. leaf extract is presented in Table 5.

Table 5. Tannic acid equivalent tannin levels in the *L. camara* Linn. leaf extract

Sample	Y	a	b	v (L)	p	m (g)	$Q_{TAET}$
(mg/g)							
1	0.124	0.00921	0.0890	0.001	10	0.1	0.3800217
2	0.125	0.00921	0.0890	0.001	10	0.1	0.3908795
3	0.125	0.00921	0.0890	0.001	10	0.1	0.3908795
4	0.124	0.00921	0.0890	0.001	10	0.1	0.3800217
5	0.125	0.00921	0.0890	0.001	10	0.1	0.3908795
6	0.123	0.00921	0.0890	0.001	10	0.1	0.3691640
Mean							0.384
SD							0.009

Abbreviation: Y=absorbance in wave length ( $\lambda$ ) of 740 nm; a=coefficient; b=constant; v=volume (liter); p=dilution; m=sample weight (gram);  $Q_{TAET}$ =tannic acid equivalent tannin levels; mg/L=milligrams per liter; mg/g=milligrams per gram; SD=standard of deviation.

The phytochemical profile of *L. camara* Linn. leaf extract is presented in Figure 7.

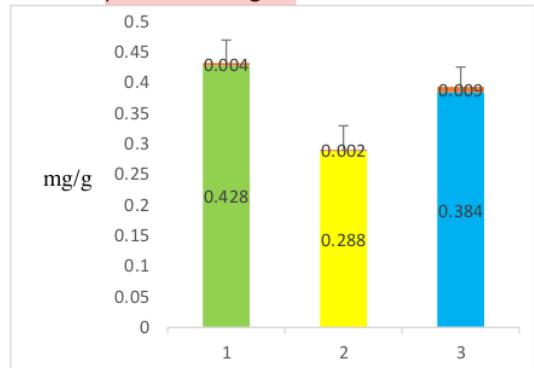


Figure 7. Phytochemical profile of *L. camara* Linn. leaf extract. 1=QEF (quercetin equivalent flavonoid). 2=GAEP (phenolic equivalent gallic acid). 3=TAET (tannin equivalent tannic acid).

#### 4. Discussion

The results of organoleptic tests on *L. camara* Linn. leaf extract in this time similar with the results of our previous research, including the form is semi-solid, the smell is similar with the smell of *L. camara* Linn. leaves, and the color is slightly blackish green [16]. The pH of *L. camara* Linn. leaf extract is normal, as it is in the range values of 4.5-6.5. The pH of *L. camara* Linn. leaf extract consistent with pH of human skin [29]. Compared to the topical formula, the pH of *L. camara* Linn. leaf extract in this study was in accordance with the pH of topical preparations containing ibuprofen [30].

The content of flavonoids, phenols, and tannins in *L. camara* Linn. leaf extract in this study was the same as the results of previous studies [20, 31, 32]. Flavonoid content in the leaves of *L. camara* Linn. also shown with extraction using acetone [33]. In addition, methanol extraction of *L. camara* Linn. leaves also showed flavonoid content [32]. In addition, the extract drying method of *L. camara* Linn. leaves also shows flavonoids and tannins [34]. The results of another study demonstrated that the leaves of *L. camara* Linn. extracted using petroleum ether (40 °C), chloroform, and methanol also contain flavonoids, and tannins [35].

QEF levels of *L. camara* Linn. leaf extract in this study were lower than the results of other studies [31, 32, 36, 37]. The results of previous studies demonstrated that various varieties of *L. camara* Linn. have QEF content ranging from 16.14±0.21 to 25.22±2.59 mg/g extract [36]. The results of another study showed that the content of QEF in dry extract of *L. camara* Linn. 12.44±2.85 mg/g [37]. Another study showed that the methanol extract of *L. camara* Linn. leaves contain QEF 243.89 mg/g extract [31]. The results of another study demonstrated that several fractions of methanol extract of *L. camara* Linn. leaves contained QEF ranging from 19.85-97.56 mg/g samples [32]. The results of other studies also revealed that the QEF content of methanol extract of aerial parts of *L. camara* Linn. from Nepal ranged from 1.87±0.16 mg/g extract [38]. On the other hands, the results of other studies demonstrate that ethanol extract of *L. camara* Linn. leaves contain low QEF, which is 0.2423±0.0068 mg/g extract [39]. These results are lower than the QEF content in our study.

GAEP levels of *L. camara* Linn. leaf extract in this study were lower than the results of other studies [31, 32, 36, 37, 38]. Previous research demonstrated that various varieties of *L. camara* Linn. have GAEP content ranging from 55.57±2.82 to 232.9±15.97 mg/g extract [36]. Other research results also showed that dry extract of *L. camara* Linn. contains GAEP 144.7±1.34 mg/g [37]. Another study showed that the methanol extract of *L. camara* Linn. leaves contain

GAEP  $563.57 \pm 2.49$  mg/g extract, while the GAEP content in flower extract  $614.79 \pm 1.54$  mg/g extract [31]. The results of another study demonstrated that the GAEP content of *L. camara* Linn. leaf extract  $10.20 \pm 0.343$  mg/g extract [38]. The results of another study demonstrated the GAEP content in various fractions of methanol leaf extract of *L. camara* Linn. ranging from  $20.25 \pm 0.41$  to  $98.81 \pm 0.27$  mg/g sample [32]. As a reference, the results of research on the content of GAEP in other plants turned out to vary, for example *Ageratina adenophora* contains GAEP  $4.70 \pm 0.059$  mg/g extract, while *Cupressus sempervirens* contains GAEP  $4.31 \pm 0.147$  mg/g extract [38].

TAET levels of *L. camara* Linn. leaf extract in [15] study were lower than the results of other studies. The results of the study demonstrated that the tannin content in [15] *L. camara* Linn. leaf extract  $98.40 \pm 6.88$  mg/g [40]. The results of another study demonstrated that the tannin content of *L. camara* Linn. extract  $0.860 \pm 0.038$  mg/g [41]. On the other hand, there are research results that demonstrate that ethanol extract of *L. camara* Linn. leaves contain low tannins, namely  $0.2179 \pm 0.0056$  mg/g extract [39]. These results were lower than the tannin content in our study. There are [8] so studies demonstrating that tannin levels from methanol extract of *L. camara* Linn. collected from a semi-arid region of Brazil is not detected [42, 43].

It is noteworthy that there are research results demonstrate that the content of GAEP and QEF in *L. rhodesiensis* extract is highest in the leaves, then the stem, while the least is found in the roots [44]. Other research results to note are about estimation of phenolics, flavonoids and tannin contents in various solvent extracts of coconut. The results showed that methanol fraction contained a total phenolic equivalent of gallic acid  $822.60 \pm 16.36$  mg/g sample, a flavonoid equivalent of quercetin  $103.30 \pm 9.78$  mg/g sample, and tannic acid equivalent tannin  $663.50 \pm 19.26$  mg/g sample [45].

Based on the data above, there are variations in QEF, GAEP, and TAET levels that are influenced by variations in plants, environment, and solvents used for extraction. Our statement is reinforced by research results showing that extraction conditions affect flavonoid levels [46]. Based on the results of these study as well as the results of other studies [47], flavonoid content was measured in plant extracts [48, 49], and herbal preparations [50, 51, 52, 53].

The limitations of this study include not examining the mineral content which can affect the levels of active substances in *L. camara* Linn. leaf extract. Therefore, we suggest for research that it is necessary to measure mineral levels, especially Fe and Zn which are related to the levels of substances in *L.*

*camara* Linn. leaf extract. The levels of these two minerals have been shown to affect the stability of QEF levels in *L. camara* Linn leaf extract cream [16].

## 5. Conclusion

The ethanolic extract of *L. camara* Linn. contains levels of QEF, GAEP, and TAET as well as  $0.428 \pm 0.004$  mg/g extract,  $0.288 \pm 0.002$  mg/g extract, and  $0.384 \pm 0.009$  mg/g extract, respectively. The content of active substance levels can be used as a reference to explore *L. camara* Linn. as a source of quercetin, gallic acid, and tannic acid.

Based on the results of our research, as well as the results of other [16] studies, and we have described above, it turns out that the leaf extract of *L. camara* Linn. contains QEF, GAEP, and TAET, but levels vary. Variation of QEF, GAEP, and TAET levels in *L. camara* Linn. leaf extract. This is influenced by the type and place of life. Nonetheless, we hope that the variations in QEF, GAEP, and TAET levels in the *L. camara* Linn. can be used as an option in the exploration and utilization of *L. camara* Linn. Thus *L. camara* Linn. not only considered as a wild plant that endangers the environment, but can be used as a source of QEF, GAEP, and TAET exploration.

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## Conflict of Interest

No conflict of interest.

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PAGE 2

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PAGE 3

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PAGE 4

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PAGE 5

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PAGE 6

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PAGE 7

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PAGE 8

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PAGE 9

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PAGE 10

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PAGE 11

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PAGE 12

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PAGE 13

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PAGE 14

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PAGE 15

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